

Fig. 1. Effect of temperature on the activity of CAT in *M. separata* acclimated at 17°C and 24°C. The data are expressed as mean ± SE. Different letters indicate significant differences (p < 0.05).

ROS levels, and the activity of antioxidant enzymes (SOD, CAT, POX, GST) and the level of malondialdehyde (MDA) in *M. separata* acclimated at 17°C and 24°C. The results showed that the activity of SOD, CAT, POX, and GST increased significantly (p < 0.05) in *M. separata* acclimated at 5°C compared to 25°C, 10°C, 15°C, 20°C, 30°C, 35°C, and 40°C. The activity of SOD, CAT, POX, and GST also increased significantly (p < 0.05) in *M. separata* acclimated at 30°C compared to 25°C, 10°C, 15°C, 20°C, 35°C, and 40°C.

The activity of SOD, CAT, POX, and GST increased significantly (p < 0.05) in *M. separata* acclimated at 5°C compared to 25°C, 10°C, 15°C, 20°C, 30°C, 35°C, and 40°C. The activity of SOD, CAT, POX, and GST also increased significantly (p < 0.05) in *M. separata* acclimated at 30°C compared to 25°C, 10°C, 15°C, 20°C, 35°C, and 40°C. The activity of SOD, CAT, POX, and GST also increased significantly (p < 0.05) in *M. separata* acclimated at 30°C compared to 25°C, 10°C, 15°C, 20°C, 35°C, and 40°C.

The activity of SOD, CAT, POX, and GST increased significantly (p < 0.05) in *M. separata* acclimated at 5°C compared to 25°C, 10°C, 15°C, 20°C, 30°C, 35°C, and 40°C. The activity of SOD, CAT, POX, and GST also increased significantly (p < 0.05) in *M. separata* acclimated at 30°C compared to 25°C, 10°C, 15°C, 20°C, 35°C, and 40°C.

The activity of SOD, CAT, POX, and GST increased significantly (p < 0.05) in *M. separata* acclimated at 5°C compared to 25°C, 10°C, 15°C, 20°C, 30°C, 35°C, and 40°C. The activity of SOD, CAT, POX, and GST also increased significantly (p < 0.05) in *M. separata* acclimated at 30°C compared to 25°C, 10°C, 15°C, 20°C, 35°C, and 40°C. The activity of SOD, CAT, POX, and GST also increased significantly (p < 0.05) in *M. separata* acclimated at 30°C compared to 25°C, 10°C, 15°C, 20°C, 35°C, and 40°C.

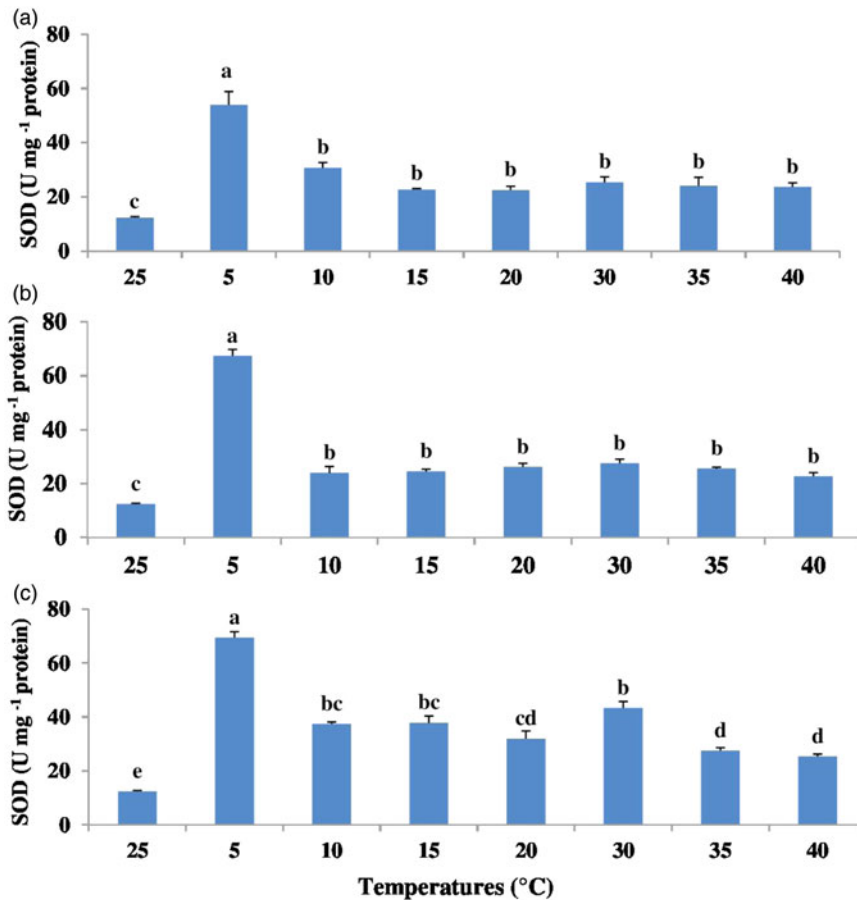


Figure 2. Effect of temperature on the SOD activity of *M. separata* at different days (1, 4 and 7 days) under different temperatures (25, 5, 10, 15, 20, 30, 35 and 40 °C). Data are expressed as mean ± SE. Error bars represent standard error (SE). Letters above bars indicate statistical significance (P < 0.05) determined by ANOVA. T = 1 day.

The effect of temperature on the SOD activity of *M. separata* (Jain & L., 1997; Xie et al., 1998) was studied. The SOD activity was measured at different temperatures (25, 5, 10, 15, 20, 30, 35 and 40 °C) for 1, 4 and 7 days. The results showed that the SOD activity was significantly higher at 5 °C (a) compared to other temperatures (b, c, d, e, f, g, h, i, j, k, l, m, n, o, p, q, r, s, t, u, v, w, x, y, z). The SOD activity was also significantly higher at 1 day (a) compared to other days (b, c, d, e, f, g, h, i, j, k, l, m, n, o, p, q, r, s, t, u, v, w, x, y, z).

Statistical analysis was performed using Duncan's multiple range test. The results are presented in Table 2. The SOD activity was significantly higher at 5 °C (a) compared to other temperatures (b, c, d, e, f, g, h, i, j, k, l, m, n, o, p, q, r, s, t, u, v, w, x, y, z).

Thermal stress

The effect of thermal stress on the SOD activity of *M. separata* was studied. The SOD activity was measured at different temperatures (25, 5, 10, 15, 20, 30, 35 and 40 °C) for 1, 4 and 7 days. The results showed that the SOD activity was significantly higher at 5 °C (a) compared to other temperatures (b, c, d, e, f, g, h, i, j, k, l, m, n, o, p, q, r, s, t, u, v, w, x, y, z).

Materials and methods

Insects

The larvae of *M. separata* were reared on a standard diet under laboratory conditions. The larvae were reared at different temperatures (25, 5, 10, 15, 20, 30, 35 and 40 °C) for 1, 4 and 7 days.

Enzyme extraction

The SOD activity was measured using a spectrophotometer. The SOD activity was measured at different temperatures (25, 5, 10, 15, 20, 30, 35 and 40 °C) for 1, 4 and 7 days.

ee eed 0.9% ae a a a 1:9
(W_e:V_a a e). H e a e ee ce ed a

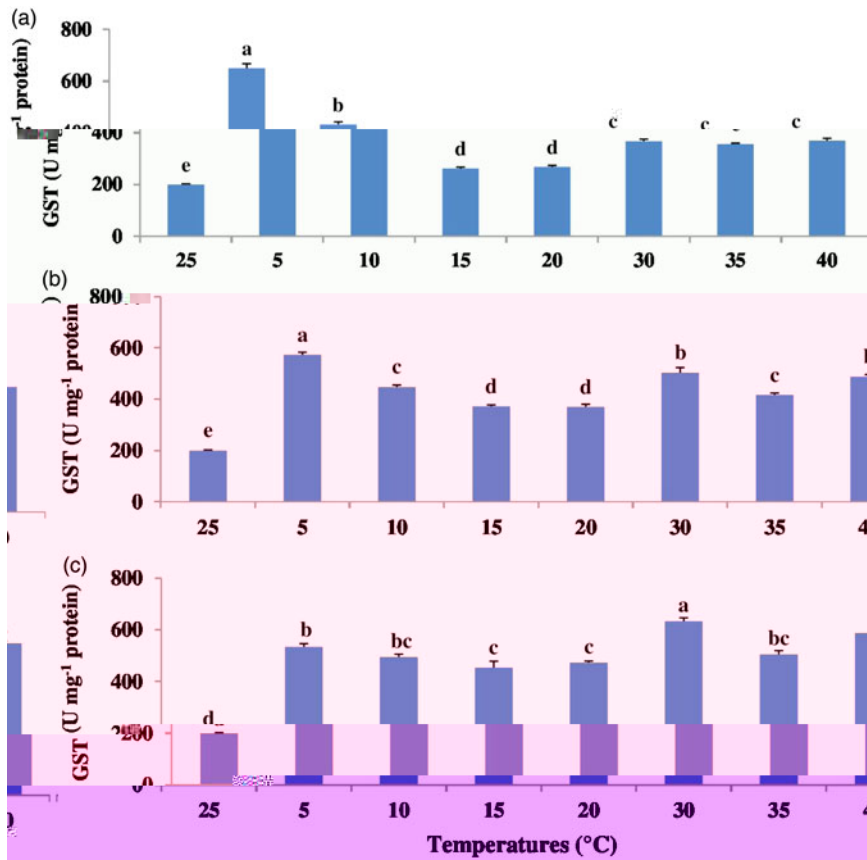


Fig. 4. Effect of temperature on GST activity in *M. separata* larvae. Data are expressed as mean ± SE. Letters above bars indicate significant differences ($P < 0.05$) determined by ANOVA. T = temperature.

de ed a e GST ac y a d e e ed a U⁻¹ e .

Statistical analysis

T e a e e ec (e e a e a d d a) e e b - e c e d e - a y a a y a a ce (ANOVA) e e e a e a de ced e SPSS 16.0 (SPSS, C ca , IL, USA); e ca e ec e e de ed, e a d e e ce e e a a ed b y T e y e , $P < 0.05$ c de ed a ca y ca .

Results

A 45 C, a ad d ed, e a d e e d a e e a e e a e .

Antioxidant enzymes

CAT ac y M. separata ad a ca y - c e a e d a b a d , c a e d e c , e - e a e ($P < 0.01$), a e a e a d a ($P < 0.01$), a d e e ac be e e e a e a d d a a ca ($P < 0.01$). Ma CAT ac y a e e e 167.67, 146.94 a d 135.50 U⁻¹ e ec ded de c d e (5 C) 1, 4 a d 7 , e ec e y (1).

SOD ac y a ca y a e d a b a d , c a e d e c , e e a e M. separata ad ($P < 0.01$), a d a e e ($P < 0.01$), a d e e a a ca e ac be e e e a e a d d - a ($P < 0.01$). T e e SOD ac y e e (53.92, 67.41 a d 69.45 U⁻¹ e) e e b e ed de c d e (5 C) 1, 4 a d 7 , e ec e y (2).

POX ac y M. separata ad a a ca y a - e c e d a a e e a e ($P < 0.01$) a d a d a ($P < 0.01$), a d e e a e d d a e a c e d - ca y ($P < 0.01$). POX ac y c e a e d ca y de - e e a e e (e e a e a 30 40 C) a 1 a d 4 , e a e c d e a d c (25 C) c d ; e e , a e 7 , e a ca e e a

POX ac y a b e e d a e e a e 15, 20, 30, 35 a d 40 C, e a e a a 10 C, ca d e e ce e e b e e d a e e e e a e c a e d e c (25 C) (3).

S ca c e a e GST ac y M. separata ad a b a d e e e a e e e b e e d a a e e a - e ($P < 0.01$) a d d a ($P < 0.01$), c a e d e c - (25 C), a d e e a a ca e ac be e e e e a e a d d a ($P < 0.01$). T e e a e GST ac y ec ded e e 649.71 a d 572.50 U⁻¹ e a 5 C 1 a d 4 , e ec e y. I add , a e 7 a 30 C GST ac y a 633.15 U⁻¹ e (4).

Total antioxidant capacity (T-AOC)

Significant differences were observed in T-AOC of *M. separata* after 1, 4, and 7 days of storage at 10, 20, 30, and 40 °C (P < 0.01). T-AOC decreased significantly (P < 0.01) during storage at 10, 20, 30, and 40 °C. The decrease in T-AOC was more pronounced at 40 °C compared to 10 °C. The decrease in T-AOC was also more pronounced at 7 days compared to 1 day of storage. The decrease in T-AOC was also more pronounced at 40 °C compared to 10 °C. The decrease in T-AOC was also more pronounced at 7 days compared to 1 day of storage.

Discussion

The present study investigated the effect of storage temperature and time on the antioxidant capacity of *M. separata*. The results showed that the antioxidant capacity of *M. separata* decreased significantly during storage at 10, 20, 30, and 40 °C. The decrease in antioxidant capacity was more pronounced at 40 °C compared to 10 °C. The decrease in antioxidant capacity was also more pronounced at 7 days compared to 1 day of storage. The decrease in antioxidant capacity was also more pronounced at 40 °C compared to 10 °C. The decrease in antioxidant capacity was also more pronounced at 7 days compared to 1 day of storage.

Significant differences were observed in ROS, CAT, and POX activity of *M. separata* after 1, 4, and 7 days of storage at 10, 20, 30, and 40 °C (P < 0.01). ROS, CAT, and POX activity decreased significantly (P < 0.01) during storage at 10, 20, 30, and 40 °C. The decrease in ROS, CAT, and POX activity was more pronounced at 40 °C compared to 10 °C. The decrease in ROS, CAT, and POX activity was also more pronounced at 7 days compared to 1 day of storage. The decrease in ROS, CAT, and POX activity was also more pronounced at 40 °C compared to 10 °C. The decrease in ROS, CAT, and POX activity was also more pronounced at 7 days compared to 1 day of storage.

e a e . S a e e e e ed b' McC d & F d c (1969) a d J a et al. (2011). SOD a d CAT ca d ec 'y e e e ce ROS ac d a ed a e . SOD e e O₂⁻ e ce d a O₂ a d H₂O₂, a d H₂O₂ e e e a 'y ed ced H₂O a d O₂ b' CAT (Ka a et al., 1997). T e b e ed e e e CAT, e a e e SOD, d' d ca e a, de e - a e , H₂O₂ a 'y e ed b' ce e e a SOD ac 'y . GST ca e ab e d e da d c e e POX, c a b e a d H₂O₂ (J a et al., 2011). I e e e d' POX ac 'y c e a ed ca 'y a e e a e (a 30 40 C) 1 a d 4 , c a ed c . S a d e e e ed b' Z a et al. (2014) e e da 'y e, *Neoseiulus cucumeris*. O e de a e a POX ac 'y a e ed 'y d ced b' e a e M. separata ad , c c - e e d a a d' *Helicoverpa armigera* (Me et al., 2009). H e e , a e e e d a (7) e a e , a ca dec e a e POX ac 'y a b e ed e e a 'y B. dorsalis (J a et al., 2011) a d e da 'y e, *N. cucumeris* (Z a et al., 2014). I c a , e d ca e ca c a e POX ac 'y a e e e e a e c a e d - a (7), a e e e ed b' Ya et al. (2010). T e e e a POX ac 'y a e e e a e d ca e a a a ed b' ca e ROS M. separata. GST a e a c a d e ce 'y e , c ca a 'y e ec a a e ab ad ec - e d e a d e b cc d de - ca , ec da e da a e , e a a d e ce a a a (B a d & Me , 2013). T e e e - 'y e a e ed e ac a c d e da - d c c e a ed b' da e e da a e . I e e e d' , e b e a ca 'y e e a e e e GST de e e a e e e a e 'y e ec M. separata ad da e da a e de e e c d . S a a da e e e a e b e e e - ed P. japonica (Z a et al., 2015), A. mylitta (J e a et al., 2013), B. dorsalis (J a et al., 2011) a d P. citri (Ya et al., 2010). T-AOC de 'y e da a a e ed , a d a a e e e a e e a e e a a da ca ac 'y e - a a (Me et al., 2009; Ya et al., 2010; Sa d a a et al., 2011). T-AOC a a e ed ca 'y e M. separata ad e e e ed e e a e (a 30 40 C) 1 a d (40 C) 4 , c a ed c . T e e da a e a T-AOC ada de a da e e a d ee ad ca a a d a ec - e e e e ed b' Z a et al. (2015), Z a et al. (2014) a d J a et al. (2011). H e e , ca d e e ce a b e ed c a ed c a e e a e e e e d a (7). A a e a e - ed b' J a et al. (2011) B. dorsalis de e a e c d . A da e e e a a ed b' a da e - 'y e ; e e , e -e 'y a c b a ce , e . e a - e (Ma d et al., 2010) a d a E (a- c e) (Ka et al., 2009) a c b e ce . A e ce d' y a c ed e e e e a c e , a a da e 'y e , e e e ROS da a e (R a et al., 2012). T e c e a e T-AOC 'y a e e a e d ca e a M. separata e 'y a da e 'y e ,

b a e de e ce ec a , c ba e a e a d e ab e a e a (J a et al., 2011).

Conclusion

O da e e ca b e e e a ed e e e a ac - d b e b a a ce ed e ac a a . I M. separata, e a e e a ca d da e ac e d c da e e . I e e e a e , a da e 'y e a e e a e da a de e ce ec a a e e a ce a da a e . T e e 'y e SOD, CAT a d GST de ca c e a e ac 'y e e e a e M. separata, a d a 'y b e ed e a a e e da e da a e d ced b' ROS. I deed, e e a a c e a ed d c ROS a e e e a e ; e e e , e e c a a 'y e ec 'y -

- Dubovskiy, I., Martemyanov, V., Vorontsova, Y., Rantala, M., Gryanova, E. & Glupov, V. (2008) Ecotoxicology and Environmental Chemistry of the Insect *Galleria mellonella* L. (Lepidoptera, Pyralidae). *Comparative Biochemistry and Physiology C: Toxicology & Pharmacology* **148**, 1–5.
- Felton, G.W. & Summers, C.B. (1995) Amino acid metabolism. *Archives of Insect Biochemistry and Physiology* **29**, 187–197.
- Ghiselli, A., Serafini, M., Natella, F. & Scaccini, C. (2000) The role of reactive oxygen species in the development of *Galleria mellonella*. *Free Radical Biology and Medicine* **29**, 1106–1114.
- Green, D.R. & Reed, J.C. (1998) Molecular cloning and characterization of the *Mythimna separata* polyphagous gene. *Science* **281**, 1309.
- Howe, G.A. & Schillmiller, A.L. (2002) Olfaction and host plant selection. *Current Opinion in Plant Biology* **5**, 230–236.
- Jena, K., Kar, P. K., Kausar, Z. & Babu, C.S. (2013) Ecotoxicology of the insecticide chlorpyrifos on the silkworm *Bombyx mori* L. (Lepidoptera: Antheraea mylitta). *Journal of Thermal Biology* **38**, 199–204.
- Jia, F.X., Dou, W., Hu, F. & Wang, J.J. (2011) Ecotoxicology of the insecticide chlorpyrifos on the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). *Florida Entomologist* **94**, 956–963.
- Jiang, X. (2004) The effect of chlorpyrifos on the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). *Acta Entomologica Sinica* **47**, 1–5.
- Jiang, X.F. & Luo, L.Z. (1997) Insecticide resistance in the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). *Acta Entomologica Sinica* **40**, 274–280. In: X.F. Jiang, X.F. & Hu, T. (Eds) *Ecological Research Sustainable Development*. Beijing, China: China Environmental Science Press.
- Jiang, X.F., Luo, L.Z. & Hu, Y. (2000) Insecticide resistance in the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). *Acta Ecologica Sinica* **20**, 288–292.
- Jiang, X., Luo, L., Zhang, L., Sappington, T.W. & Hu, Y. (2011) Resistance to chlorpyrifos in the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). *Environmental Entomology* **40**, 516–533.
- Jing, X.H., Wang, X.H. & Kang, L. (2005) Chlorpyrifos resistance in the locust *Locusta migratoria* (Orthoptera: Acrididae). *Insect Science* **12**, 171–178.
- Joanisse, D. & Storey, K. (1996) Olfaction and host plant selection in the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). *Journal of Experimental Biology* **199**, 1483–1491.
- Joanisse, D.R. & Storey, K.B. (1998) Olfaction and host plant selection in the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). *Insect Biochemistry and Molecular Biology* **28**, 23–30.
- Kashiwagi, A., Kashiwagi, K., Takase, M., Hanada, H. & Nakamura, M. (1997) Chlorpyrifos resistance in the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). *Acta Entomologica Sinica* **40**, 274–280.

NADP+ and NADH/NAD+ in *Drosophila melanogaster*.
Mechanisms of Ageing and Development **56**, 223–235.

Wang, Y., Oberley, L.W. & Murhammer, D.W. (2001) A study of the role of reactive oxygen species in aging. *Free Radical Biology and Medicine* **30**, 1254–1262.

Wang, G.P., Zhang, Q.W., Ye, Z.H. & Luo, L.Z. (2006) The effect of the insecticide Mythimna separata (Lepidoptera: Noctuidae) on the host. *Bulletin of Entomological Research* **96**, 445–455.

Xinfu, J., Yueqiu, L. & Lihi, L. (1998) Effect of the insecticide Mythimna separata Walker. *Journal of Beijing Agricultural College (China)* **13**, 20–26.

Yang, L.H., Huang, H. & Wang, J.J. (2010) A study of the effect of the insecticide Mythimna separata Walker on the host. *Journal of Beijing Agricultural College (China)* **25**, 1–5.