

ARTICLE

Knockdown of TOR causing ovarian diapause in a genetically stable brachypterous strain of *Nilaparvata lugens* gen

Fangzhou Liu¹  | Kaiyin Li¹ | Wanlun Cai¹ | Jing Zhao¹ | Yulan Zou² | Hongxia Hua¹

¹Henan Provincial Key Laboratory of Insect Molecular Biology, Henan University of Science and Technology, Zhengzhou, China
²Henan Provincial Key Laboratory of Insect Molecular Biology, Henan University of Science and Technology, Zhengzhou, China

Correspondence

Hongxia Hua, Key Laboratory of Insect Molecular Biology, Henan University of Science and Technology, Zhengzhou 450007, China.
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Abstract

Brachypterous (BPH), *Nilaparvata lugens* (Stål) (Homoptera: Delphacidae), is a major pest of rice. BPH is a diapausing insect during winter. The diapause is a survival strategy for overwintering. TOR signaling pathway is involved in the regulation of diapause. In this study, we investigated the effect of TOR knockdown on the diapause of BPH. The results showed that TOR knockdown significantly reduced the diapause rate of BPH. This suggests that TOR signaling pathway is involved in the regulation of diapause in BPH.

























ii  ii  , i i  a i  i  BPH.A ii  ii i  a
i MF, i a  ii  ii i  a  i BF(J  ,1963;Kii  ,1956).
l abia i a ab , BPH i BF i  i a  i a i  a i (S b ,
Mi ia,Ya a  iMa a i&Ma a,2002); abia  ii , BPH i MF
i abii a  iia i a i (Kii  ,1956).
l a ii  i ,MF a a a i ii i a BF a (Kii  ,1965).
T i i  i  a b  i  a a i a a , a a i a
i i a i (C  ,C  ,Ya  ,&Yi,1979).BPH i i i b *Insulin Receptor 1 a Insulin
Receptor 2*

TABLE 1 | List of primers

GenBank accession number	Name of primer	Primer sequences	Length of PCR product
JQ793898.1	NITOR-F	GATCGGCATGAGGGAGGGAGACA	6,768 b
	NITOR-R	CGACGACGGTACTACTGCGTTTGG	
	1NITOR-F	TAATACGACTCACTATAGGACCAG TGAAATGCTCGTAAACA	550 b
	1NITOR-R	TAATACGACTCACTATAGGCCAGGTGCAGG TAATCGTCCAG	
	2NITOR-F	TAATACGACTCACTATAGTTGACGG TCACTCACTACTGCA	492 b
	2NITOR-R	TAATACGACTCACTATAGGGCTCTTTGTT TCGTCCCATACC	
U76561	GFP-F	TAATACGACTCACTATAGGGTAA CGGCCACAAGTTCAG	400 b
	GFP-R	TAATACGACTCACTATAGGTCCGG CATGATATAGACGTT	
EU179846.1	PCR-NIActin1-F	CCAACCGTGAGAAGATGACC	256 b
	PCR-NIActin1-R	GATGTCACGCACGATTTTAC	
JQ793898.1	PCR-NITOR-F	AACCCATGGAGGTGACAGG	143 b
	PCR-NITOR-R	ATGAGGCGCCAGTTGAGCAG	
KY827832	PCR-NIE74B-F	AACAACATAATAGGCACAGTC	175 b
	PCR-NIE74B-R	GGAATGGCGAAGAAGTATC	
FJ263049.1	PCR-NIEcR-F	GCCAGAAAGTACGACGTGAA	234 b
	PCR-NIEcR-R	TTGGATCTTCTCCACCTTCC	
19JF345255	PCR-NIFoxA-F	GCGGAGTTATGTTGTGTGTA	193 b
	PCR-NIFoxA-R	CTGAGCCTTGTAGCATGTTGAA	
AB353856.1	PCR-NIVg-F	TCTTCATCATCTCCTCCTCTTC	173 b
	PCR-NIVg-R	TCCTGGTTGTTGCATTGTCATT	

2.3 | Synthesis of dsRNA

To synthesize dsRNA, the primers listed in Table 1 were used to amplify the target DNA fragments. The PCR products were purified and then ligated with BamHI and XbaI sites. The resulting fragments were digested with BamHI and XbaI to generate dsRNA. The dsRNA was then purified and stored at -80°C. For RNAi, the dsRNA was injected into the prothoracic coxae of 3rd instar larvae. The efficiency of RNAi was determined by measuring the mortality and the weight of the larvae. The results showed that the dsRNA targeting NITOR significantly reduced the mortality and weight of the larvae (Table 1).

2.4 | RNAi using microinjection

To perform RNAi using microinjection, the dsRNA was injected into the prothoracic coxae of 3rd instar larvae. The efficiency of RNAi was determined by measuring the mortality and the weight of the larvae. The results showed that the dsRNA targeting NITOR significantly reduced the mortality and weight of the larvae (Table 1).

GFP) a i a i a a a a a .A a 250
a i a a i a a a (i., i a 750 i a a
RNA).T a a a b b Li a.(2015).T i a BPH a a ai
a i i .A a a i , i a a a RT-PCR a a i a 3,6,7,9,11 a a
i i .

2.5 | Developmental duration and ovary observation

Ea BPH a i RNA a b a 6 i i a a i i i .N
a i i - a .T BPH ai i i i - i i
(WPI, Sa a a) a 1,3,5, a 8 a a a , i 1× a -b ai (PBS) b a -
i i 4% a i 1×PBS 20 i a a .Di a i a i
10 i i 0.2% T i -X 100 (C N.:T9824, Sa a a a, Si a) i 1×PBS. A a i , a i -
a i a O (SZX16, O ,T ,Ja a) a a P90P Di i a Ca a
(P90, Ni ,T ,Ja a).R i i BPH i a b .

2.6 | qRT-PCR

TRI a (C N.:15596018, I i ,Ca ba) a a a RNA. *Actin1* (G Ba a i
b EU179846.1) a a a i a .SYBR P i E Ta (C N.:RR420A, Ta a a, Da a ,
C i a) a RT-PCR a a i a ABI P i 7300 i (A i Bi) a i
a a ' i i .Ea ai i i a i a i a i a 100 300b PCR
(Tab 1), a PCR a i i i a i a a a .O i i
85 110% i i a a 98% a i i i RT-PCR a a i .Ea RNA a a
a i b i a i , a i a i i a a a a a
a .T bi a a a a .T a i i a a a
a i Lia a S i (2001).S i RT-PCR a i Tab 1.

2.7 | Sequence comparison and phylogenetic relationship

P i i i ba a i a i , i ,
i b i b - i i i M a E i a G A a i (MEGA) a .
T b a a a i i 1,000 a .

2.8 | Data analysis

A a a a i SPSS i 18.O - a ANOVA a a a i b a ,
a a - a ANOVA a a a i a RNA. P a a
a i b a i a a i .

3 | RESULTS

3.1 | *NITOR* expression patterns during different developmental stages in BS and MS BPH

T a *NITOR* a i b a a a i BPH; i a i a ai
i a a .D i a a , *NITOR* i a i i a i BS a
MS a i a, b i i i *NITOR* i a a a .
T *NITOR* i i BS a i i a i MS a a 1,2, a 4 a a

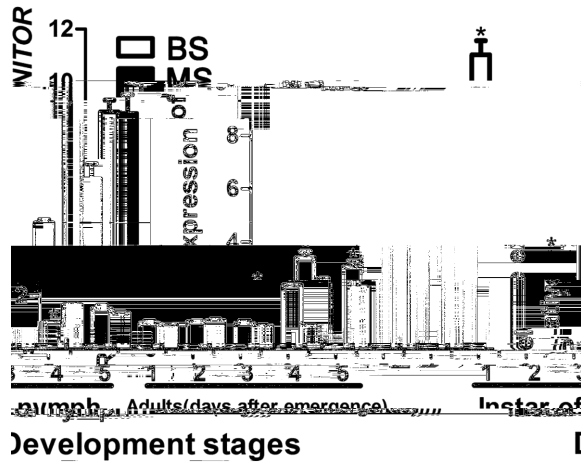


FIGURE 1 T a i i a NITOR a i a a MSa BS; BPH *actin1* a a a .T i a a i a i a a BS.T a a i a ba bi a a .E ba i a a .Ba ab i a a i i i i a b BPH MSa BS,a i i - a ANOVA ($P < 0.05$)

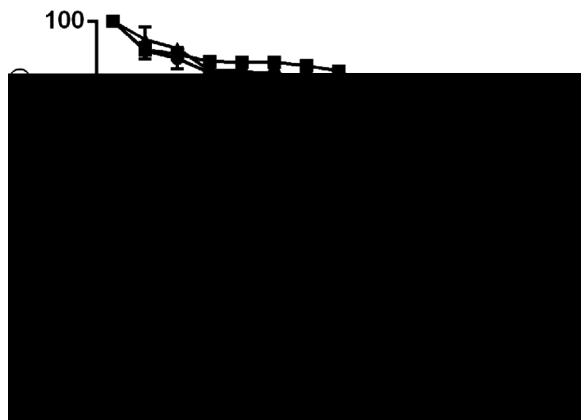


FIGURE 2 S ia a BS BPH i 1NITOR a 2NITOR i i i . GFP i i i a a .Ea i i 45 .S ia GFP, 1NITOR, a 2NITOR a a i a - a ANOVA

i i i i i i a a 5a b BSa MS(Fi 1).

3.2 | Effects of knockdown of NITOR on the survival of BS BPH

B a i i NITOR i BS a i i a i MSa a 1,2,a 4 a a , NITOR a i i BS i ia NITOR i i i a a i BPH.

A i i RNA(1NITOR, 2NITOR, a GFP) i i - i a , ia a a i NITOR a GFP i i 1 7 a a i i ($F_{2,6} = 0.538; P = 0.609; Fi 2$).I a , ia a i a i NITOR a i i a a i a i GFP 8 15 a a i i (BS: $F_{2,6} = 12.568; P = 0.007; Fi 2$).F 1 7 a a i i , a

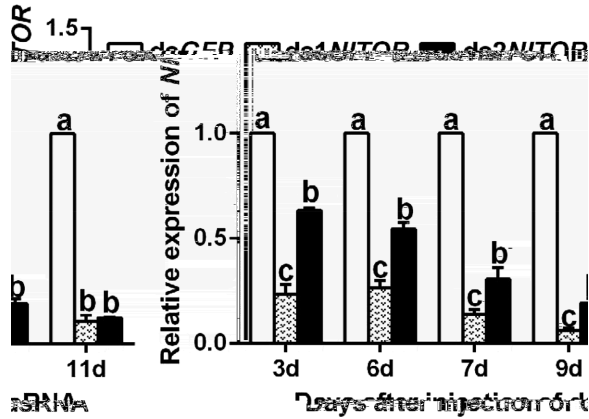


FIGURE 3 T a i i NITOR a i i RNA i i - i a i BS. E ba i a a a .T i a a i a a a i i i . Ba ab i i i a a a i i a a a a a i i - a ANOVA ($P < 0.05$). T a a i a ba bi-

BPH i a a , i 8 15 a a i i , a BPH i a a .T i i a a NITOR a a BPH a b BPH .

3.3 | Effects of dsRNA injection on NITOR gene expression in BS

I BS, i i 1NITOR a 2NITOR i i - i a i i a i NITOR RNA a a a i i .F 3 11 a a i i NITOR, a i NITOR a i i a b 36.9 94.1%(F_i 3)($P <$

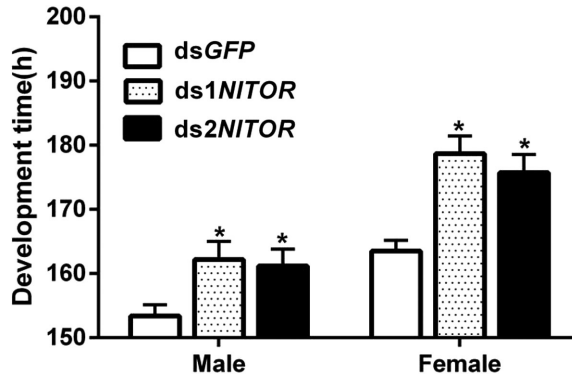


FIGURE 4 Development time of male and female *B. dorsalis* pupae under different dsRNA treatments. The development time (h) of male and female pupae was significantly different between dsGFP, ds1NITOR, and ds2NITOR treatments in both sexes (ANOVA, $P < 0.05$). Error bars represent standard deviation. Letters above bars indicate significant differences (ANOVA, $P < 0.05$).

3.6 | Knockdown of NITOR inhibited ovarian development

Western blot analysis showed that the expression of NITOR protein was significantly reduced in the ovaries of female *B. dorsalis* pupae treated with ds1NITOR or ds2NITOR compared to the dsGFP control (Figure 5). The reduction in NITOR protein levels was accompanied by a decrease in the number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae. The number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae was significantly lower than that of the dsGFP control (Figure 6). The number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae was significantly lower than that of the dsGFP control (Figure 6). The number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae was significantly lower than that of the dsGFP control (Figure 6). The number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae was significantly lower than that of the dsGFP control (Figure 6).

3.7 | Effects of in vivo knockdown of NITOR on the expression of NIFoxA and NIVg related to ovarian development

Real-time PCR analysis showed that the expression of *NIFoxA* and *NIVg* mRNA was significantly reduced in the ovaries of female *B. dorsalis* pupae treated with ds1NITOR or ds2NITOR compared to the dsGFP control (Figure 7). The reduction in *NIFoxA* and *NIVg* mRNA levels was accompanied by a decrease in the number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae. The number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae was significantly lower than that of the dsGFP control (Figure 6). The number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae was significantly lower than that of the dsGFP control (Figure 6). The number of ommatidia in the ovaries of ds1NITOR and ds2NITOR treated pupae was significantly lower than that of the dsGFP control (Figure 6).

4 | DISCUSSION

TOR signaling pathway plays a crucial role in the regulation of ovarian development in *B. dorsalis* (Ogawa et al., 2000; Zhang et al., 2000). In this study, we demonstrated that the knockdown of NITOR protein significantly inhibited ovarian development in female *B. dorsalis* pupae (Liu et al., 2016; Ma et al., 2009; Pan et al., 2006). NITOR is a key component of the TOR signaling pathway, and its knockdown leads to a decrease in the number of ommatidia in the ovaries (Liu et al., 2016; Zhang et al., 2015; Zhang et al., 2017). Downregulation of NITOR protein in the ovaries of female *B. dorsalis* pupae leads to a decrease in the number of ommatidia in the ovaries (Cao et al., 1979). Western blot analysis showed that the expression of NITOR protein was significantly reduced in the ovaries of female *B. dorsalis* pupae treated with ds1NITOR or ds2NITOR compared to the dsGFP control (Figure 5).

a BPH , BPH .W i a a NITOR a a
i a i a i i i NITOR i a a i
b i .
P i i a a NITOR a a a a i JH a i -
a a (NIJHAMT), a a i JHIII RNA i(NITOR) a a a
a b RNA i i a TOR a a i JH b i i a AA - a V i i
N. lugens (L a., 2016). O a NITOR a i i a a a .NITOR -
a i i a i NIE74Ba NIEcR.T a i i a TOR a a ,
JH b i i , a i a a a b i .
I , NITOR a i NIFoxA a NIVg.T i a -
a i Z a i a. (2015). W a i a TOR i a i i BPH i
, a i i a i i a a i i a a b a i i
NIFoxA NIVg.

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T i a b a a Na i a Na a S i F a i C i a (N .31171846).

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